

# Sulfatase Activity Assay Kit

ARG83565 Sulfatase Activity Assay Kit can be used to measure Sulfatase in Tissue extracts, Cell lysate, Cell culture media, Other biological fluids

Catalog number: ARG83565

Package: 100 tests

For research use only. Not for use in diagnostic procedures.

## TABLE OF CONTENTS

SECTION	Page
PRINCIPLE OF THE ASSAY	3
MATERIALS PROVIDED & STORAGE INFORMATION	3
MATERIALS REQUIRED BUT NOT PROVIDED	4
TECHNICAL HINTS AND PRECAUTIONS	4
SAMPLE COLLECTION & STORAGE INFORMATION	5
REAGENT PREPARATION	6
ASSAY PROCEDURE	7
CALCULATION OF RESULTS	8

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#### PRINCIPLE OF THE ASSAY

ARG83565 Sulfatase Activity Assay Kit provides a simple and sensitive method for monitoring sulfatase activity in various samples. The kit measures the hydrolysis of a sulfate ester to 4-nitrocatechol, which can be measured at a colorimetric readout at 515 nm.

#### **MATERIALS PROVIDED & STORAGE INFORMATION**

Store Positive Control and Substrate at-20 °C and protect from light, all other component at 2-8°C. Use the kit before expiration date.

Component	Quantity Storage	
Microplate	1 X 96-well plate	
Standard	1 vial (lyophilized)	4 °C
Positive Control	1 vial (lyophilized)	-20 °C
Assay Buffer	4x 30 ml	4 °C
Reaction Buffer	10 ml	4 °C
Substrate	1 vial (lyophilized)	-20 °C
Stop Solution	10 ml	4 °C

#### MATERIALS REQUIRED BUT NOT PROVIDED

- Microplate reader capable of measuring absorbance at 450 nm
- Pipettes and pipette tips
- Deionized or distilled water

#### **TECHNICAL HINTS AND PRECAUTIONS**

- Wear protective gloves, clothing, eye, and face protection especially while handling blood or body fluid samples.
- Store Positive Control and Substrate at-20 °C and protect from light, all other component at 2-8°C. Use the kit before expiration date.
- It is highly recommended that the standards and samples be assayed in at least duplicates.
- Change pipette tips between the addition of different reagent or samples.

#### SAMPLE COLLECTION & STORAGE INFORMATION

<u>Cell and bacteria</u>- Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for  $5 \times 10^6$  cell or bacteria, sonicate (with power 20%, sonicate 3s, interval 10s, repeat 30 times); centrifuged at 10,000g 4 °C for 20 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

<u>**Tissue</u>**- Weigh out 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 10,000g 4 °C for 20 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.</u>

#### **REAGENT PREPARATION**

- Substrate: Reconstitute the Substrate with 1 ml of <u>Reaction Buffer</u>. Allow the Substrate keep on bench for few minutes. Make sure the Substrate is dissolved completely and mixed thoroughly before use.
  The diluted Substrate is stable for 4 weeks at-20°C.
- Standard: Reconstitute the Standard with 1 ml of <u>distilled water</u>. Allow the Standard keep on bench for few minutes. Make sure the Standard is dissolved completely. Perform 2-fold serial dilutions of the top standards to make the standard curve.

The diluted **Standard** is stable for 4 weeks at-20°C.

 Dye Reagent A: Reconstitute the Dye Reagent A with 1 ml of <u>distilled water</u>. Allow the Dye Reagent A keep on bench for few minutes. Make sure the Dye Reagent A is dissolved completely and mixed thoroughly before use. The diluted Dye Reagent A is stable for 4 weeks at-20°C.

#### ASSAY PROCEDURE

Standards and samples should be assayed in at least duplicates.

- 1. Add **80 µl** <u>Reaction Buffer</u> into <u>all wells</u>.
- 2. Add **10 µl** <u>Sample</u> into <u>Sample</u> wells
- 3. Add **10** µl <u>Distilled water</u> into <u>Control</u>, <u>Standard</u> and <u>Blank</u> wells.
- 4. Add **10** µl <u>Positive Control</u> into <u>Positive Control</u> wells.
- 5. Add **10 µl** <u>Substrate</u> into <u>all wells</u>.
- 6. Add **10 µl** <u>Standard</u> into <u>Standard wells</u>.
- 7. Mix well incubate for **30 min** at **37 °C**
- 8. Add **10 μl** per <u>Dye Reagent</u> into <u>all wells</u>.
- 9. Mix well. Read the OD at **515nm**.

Reagent	Sample	Control	Standard	Blank	Positive Control	
Reaction Buffer	80 µl	80 µl	80 µl	80 µl	80 µl	
Sample	10 µl	-	-	-	-	
Distilled water	-	10 µl	10 µl	10 µl	-	
Positive Control	-	-	-	-	10 µl	
Substrate	10 µl	10 µl	10 µl	10 µl	10 µl	
Standard	-	-	10 µl	-	-	
Mix well incubate for 30 min at 37 °C						
Dye Reagent	100 µl	100 µl	100 µl	100 µl	100 µl	
Read the OD at 515 nm						

Summary of Sulfatase Activity Assay Kit Procedure

### **CALCULATION OF RESULTS**

Calculate the average absorbance values for each set of samples, standard and blank.

a.) Definition:

One unit of sulfatase activity is defined as the enzyme which generates 1  $\mu mol$  of 4-nitrocatechol per minute at 37°C.

C<sub>Protein</sub>: the protein concentration, mg/ml; C<sub>Standard</sub>: the concentration of Standard, 5 mmol/L = 5 μmol/ml; W: the weight of sample, g; N: the quantity of cell or bacteria, N × 104; V<sub>Standard</sub>: the volume of standard, 0.01 ml; V<sub>Sample</sub>: the volume of sample, 0.01 ml; V<sub>Assay</sub>: the volume of Assay buffer, 1 ml; T: the reaction time, 30 min.

b.) Calculation:

Formula:

a). According to the <u>Protein Concentration</u>:

Sulfatase (U/mg) = (Cstandard x Vstandard) x (ODsample - ODcontrol) / [(ODstandard - ODBlank) x (CProtein x Vsample) x T]

=0.167 × (OD<sub>Sample</sub>- OD<sub>Control</sub>) / [(OD<sub>Standard</sub>- OD<sub>Blank</sub>) × C<sub>Protein</sub>]

b). According to the weigh:

Sulfatase (U/mg) = (C<sub>Standard</sub> x V<sub>Standard</sub>) x (OD<sub>Sample</sub>- OD<sub>Control</sub>) / [(OD<sub>Standard</sub>-

OD<sub>Blank</sub>) x (V<sub>Sample</sub> x W/ V<sub>Assay</sub>) x T]

=0.167 × (OD<sub>Sample</sub>- OD<sub>Control</sub>) / [(OD<sub>Standard</sub>- OD<sub>Blank</sub>) x W]

b). According to the Cells or bacteria:

Sulfatase (U/10<sup>4</sup>) = (Cstandard x VStandard) x (ODsample- ODControl) / [(ODstandard-

OD<sub>Blank</sub>) x (V<sub>Sample</sub> x N/ V<sub>Assay</sub>) x T]

=0.167 × (OD<sub>Sample</sub>- OD<sub>Control</sub>) / [(OD<sub>Standard</sub>- OD<sub>Blank</sub>) x N]